

Application of Machine Learning to elucidate the biology predicting response in the I-SPY 2 neoadjuvant breast cancer trial

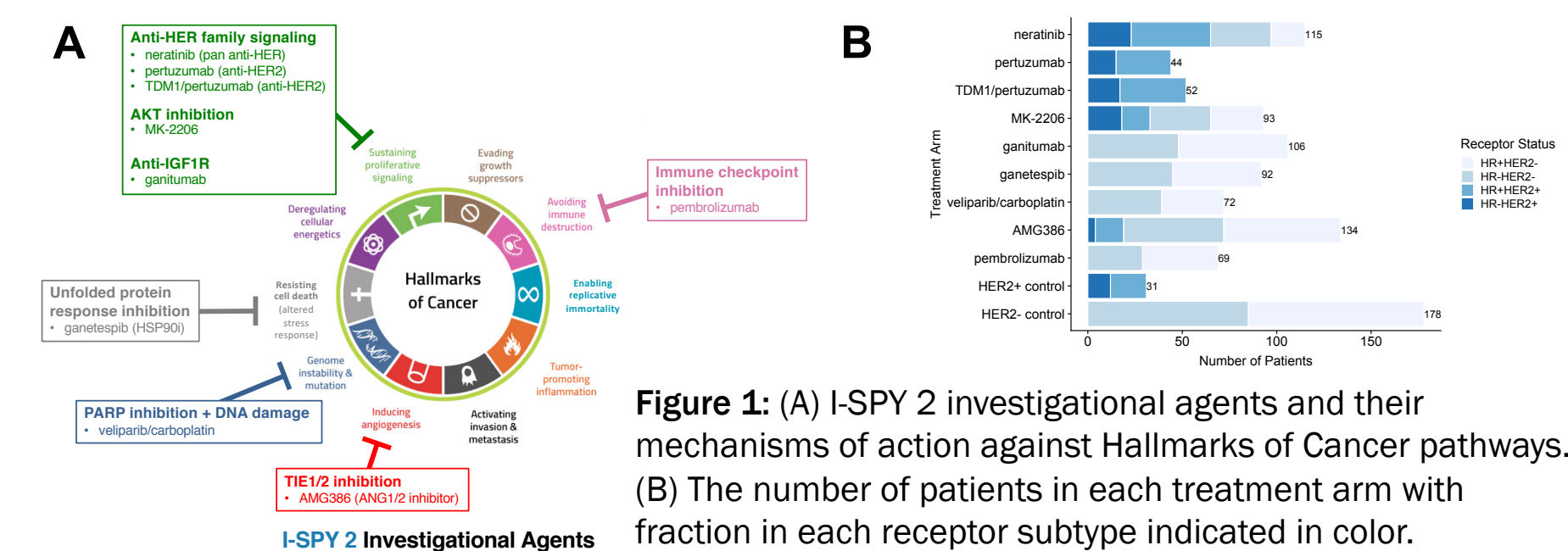
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BACKGROUND

Machine learning relies on algorithms that learn patterns in large, complex datasets to predict outcomes. The adaptive, neoadjuvant I-SPY 2 TRIAL evaluates novel agents added to standard therapy, and identifies their most responsive subtype. While previously proposed genes/signatures reflecting an agent's mechanism of action predicted pathologic complete response (pCR) in some treatment arms/subtypes, not all arms had strong predictive biomarkers. We leverage machine learning to explore the limitations of using only known mechanisms of action in predicting pCR, and the extent to which biology outside known drug action improves response prediction in the first 10 arms of the trial.

Our study involves 986 patients with pre-treatment gene expression and pCR data across 10 treatment arms including inhibitors of HER2: neratinib (N), pertuzumab (P), TDM1/pertuzumab (TDM1/P); AKT (MK-2206; M); IGF1R (ganitumab); HSP90 (ganetespib); PARP/DNA repair (veliparib/carboplatin; VC); ANG1/2 (AMG386); immune checkpoints (pembrolizumab; Pembro); and a shared control arm (Ctr) (Figure 1).



I-SPY2's ADAPTIVE TRIAL DESIGN

I-SPY 2 is a multicenter, phase 2 trial using response-adaptive randomization within biomarker subtypes to evaluate a series of novel agents when added to standard neoadjuvant therapy for women with high-risk stage II/III breast (Figure 2). Within each patient subtype, participants are assigned to one of several investigational therapies or the control regimen (4:1). Randomization probabilities are weighed by the probability of achieving a pCR within each subtype for each agent and adapts over the course of the trial. *The primary endpoint is pathologic complete response (pCR, no residual disease in breast or nodes) at surgery.*

The goal is to identify/graduate regimens that have ≥85% Bayesian predictive probability of success (statistical significance) in a 300-patient phase 3 neoadjuvant trial, defined by hormone-receptor (HR) & HER2 status & MammaPrint (MP).

Regimens may leave the trial for one of four reasons: Graduate, Drop for futility (< 10% probability of success), Drop for safety issues, or accruing maximum sample size (10% < probability of success < 85%).

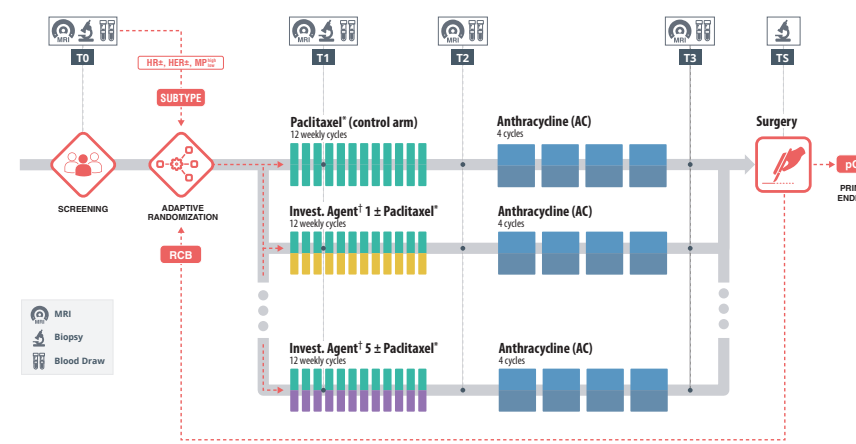


Figure 2: I-SPY 2 study schema and adaptive randomization based on probabilities of agents of achieving pCR within a given subtype.

METHODOLOGY

Each treatment arm/receptor subtype subgroup with at least 20 patients (n=19) was evaluated independently with 25% of data held out as independent test sets. Log2 transformed data was centered and scaled. We then used a 3-fold cross validation technique with 10 repeats applying different resampling methods. Random Forest ensemble algorithm was implemented with recursive feature elimination (Figure 3).

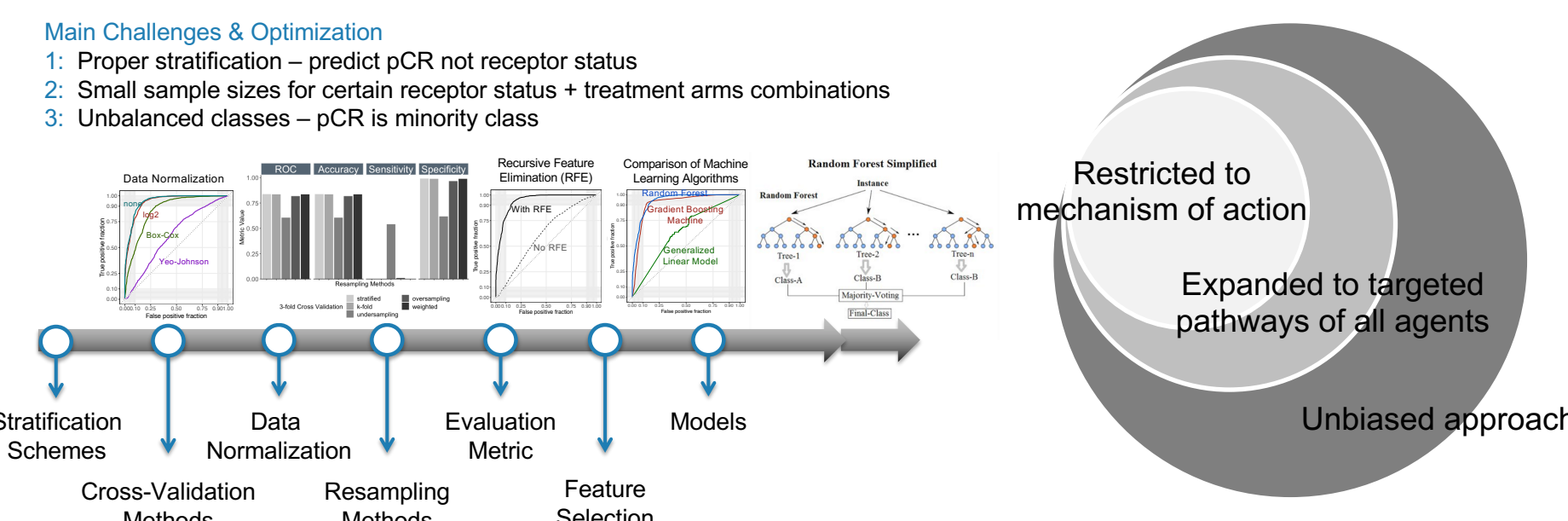


Figure 3: Schematic outlining main challenges, methodological considerations and optimization steps. (Random Forest Simplified schematic from Wikimedia Commons)

In combination with clinical data, a three-pronged feature-selection approach was employed: (1) restricted to mechanism of action genes: AKT/PI3K/HER (m=10 genes), IGF1 (m=11), HSP90 (m=88), DNA repair (m=79), TIE1/2 (m=11), and immune (m=61), as well as HER2 amplicon genes; (2) expanded to include targeted pathways for all 10 agents/combinations plus ESR1 and proliferation genes (m=339); (3) an unbiased whole genome approach (m=17,990) (Figure 4).

Models were considered predictive if AUROC ≥ 0.75, Sensitivity ≥ 0.6 and Specificity ≥ 0.6 in cross validation and independent test sets.

PREDICTION OF PATHOLOGIC COMPLETE RESPONSE

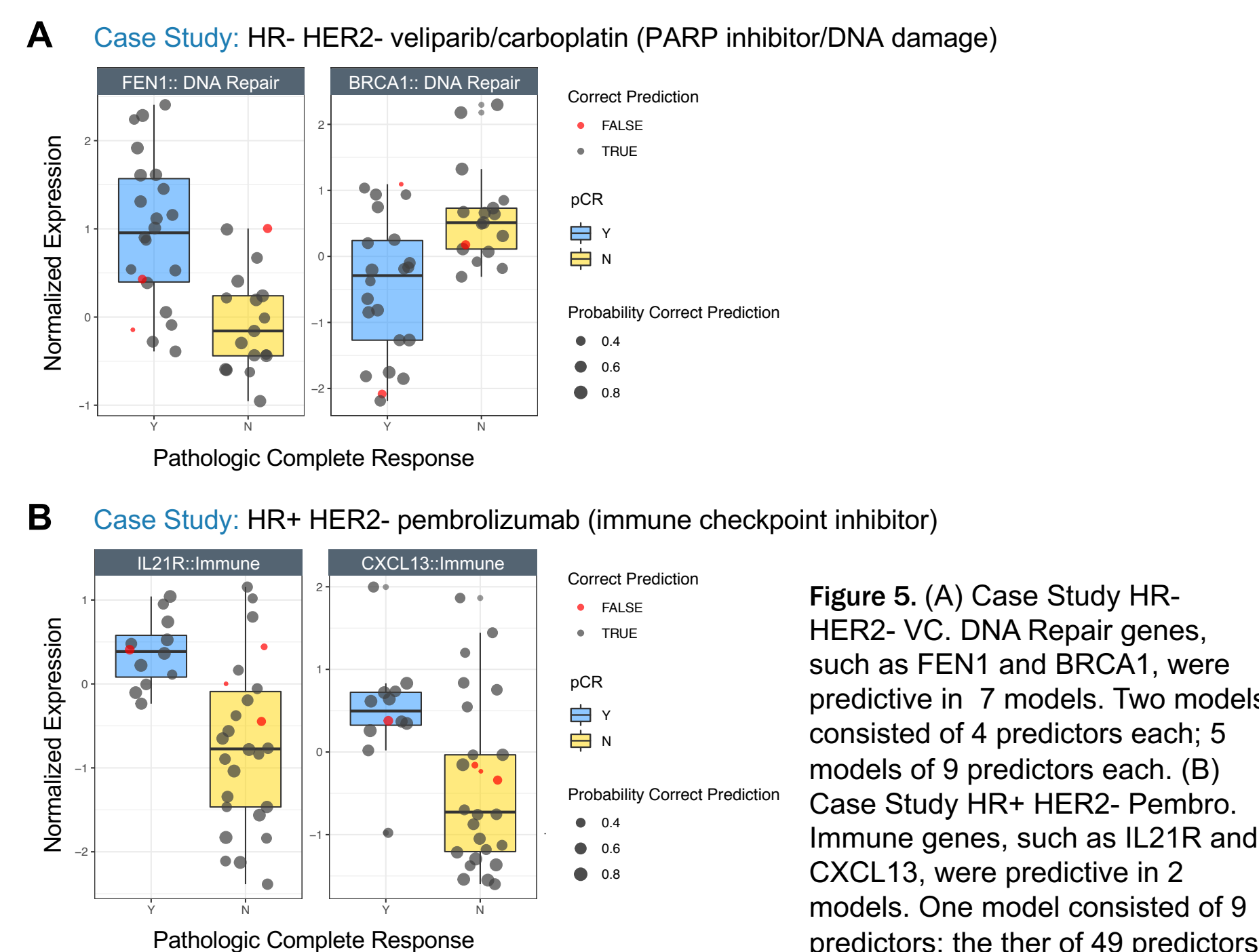
Table 1 summarizes the results of our analysis (red=Predictive model; blue=No predictive model; grey/NA=insufficient or no data). In total, we identified predictive biomarkers in 14 of 19 subgroups across the three feature selection approaches: (1) restricted to mechanism of action genes; (2) expanded to include targeted pathways for all 10 agents/combinations plus ESR1 and proliferation genes; (3) an unbiased whole genome approach.

Table 1:

| Agent | HR+HER2- | | | HR-HER2- | | | HR+HER2+ | | | HR-HER2+ | | |
|---------------------------------------------------|--------------------|---------------------|----------|--------------------|---------------------|----------|--------------------|---------------------|----------|--------------------|---------------------|----------|
| | Number of Patients | Mechanism of Action | Unbiased | Number of Patients | Mechanism of Action | Unbiased | Number of Patients | Mechanism of Action | Unbiased | Number of Patients | Mechanism of Action | Unbiased |
| neratinib (pan-anti-HER) | 34 | | | 32 | No | No | 42 | No | Yes | 23 | No | Yes |
| pertuzumab (anti-HER2) | NA | | | NA | | | 29 | Yes | Yes | 35 | No | Yes |
| TDM1/pertuzumab (anti-HER2) | NA | | | NA | | | 35 | No | No | 37 | | |
| MK-2206 (AKT inhibitor) | 26 | No | No | 32 | No | No | 34 | No | No | 36 | | |
| ganitumab (IGF1R inhibitor) | 58 | No | Yes | NA | 48 | No | NA | | | NA | | |
| ganetespib (HSP90 inhibitor) | 48 | No | Yes | 45 | No | No | NA | | | NA | | |
| veliparib/carboplatin (PARP inhibitor/DNA damage) | 33 | Yes | Yes | 39 | Yes | Yes | NA | | | NA | | |
| AMG386 (ANG1/2 inhibitor) | 62 | No | Yes | 58 | No | No | 35 | | | NA | | |
| pembrolizumab (immune checkpoint inhibitor) | 38 | Yes | Yes | 29 | No | No | NA | | | NA | | |
| HER2+ control | NA | | | NA | | | 19 | | | 12 | | |
| HER2- control | 94 | No | No | 84 | No | No | NA | | | NA | | |

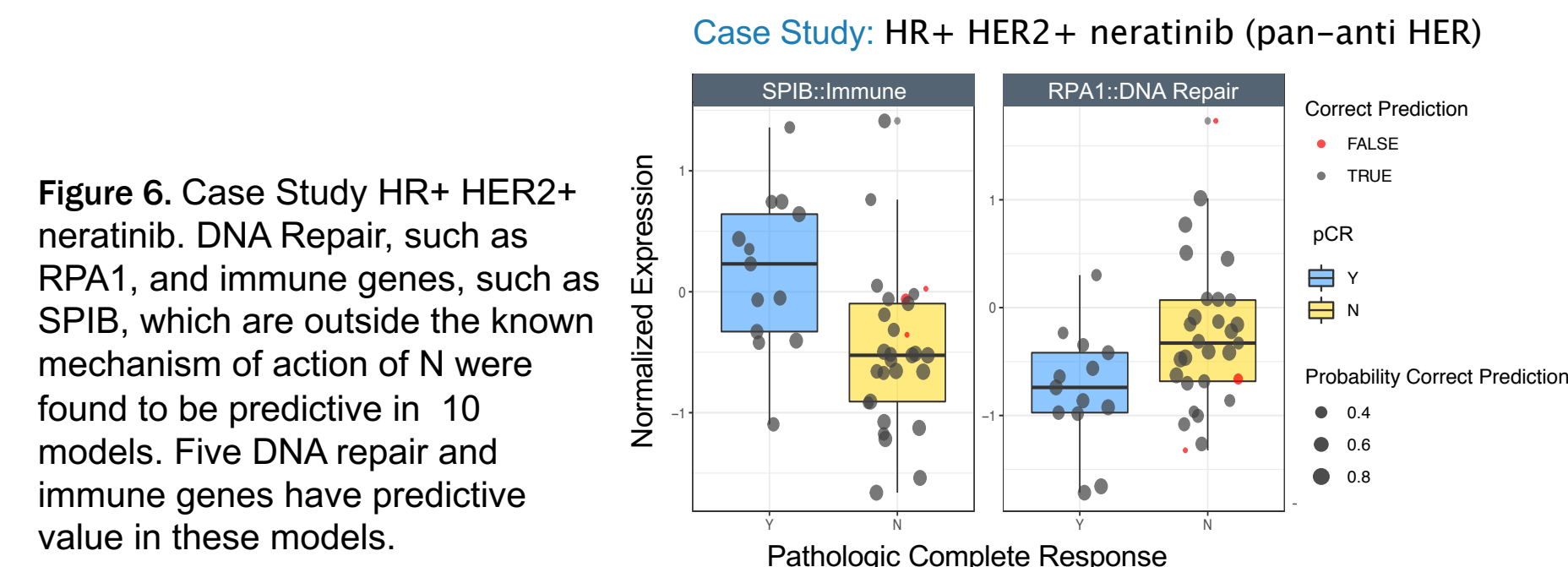
PREDICTORS WITHIN MECHANISM OF ACTION

Prediction of pCR using only genes reflecting the known mechanism of the drug succeeded in 5 subgroups, with DNA repair genes (Figure 5A) predicting VC response and immune genes (Figure 5B) predicting Pembro response in HR+HER2- and HR-HER2- subsets, and AKT/PI3K/HER + HER2 amplicon genes predicting pertuzumab response in HR+HER2+ patients.



PREDICTORS ACROSS MECHANISMS OF ACTION

Expansion of the feature set to include genes associated with all mechanisms of action of all drugs proved sufficient to produce good predictive models in 8 of 19 subgroups. Examples include DNA repair + immune genes predicting response to ganitumab in HR+HER2- and to neratinib in HR+HER2+ (Figure 6).



UNBIASED PREDICTORS

An unbiased approach using all data yielded predictive power in 8 of 19 subgroups, including 5 with no predictive models from the first two approaches. Examples include HR-HER2- neratinib predictors enriched for metabolic, cell division and membrane protein proteolytic processes; HR+HER2+ TDM1/P enriched for metabolic, stress response and cell cycle processes (Figure 7); and HR-HER2- MK-2206 predictors containing Ser/Thr kinases.

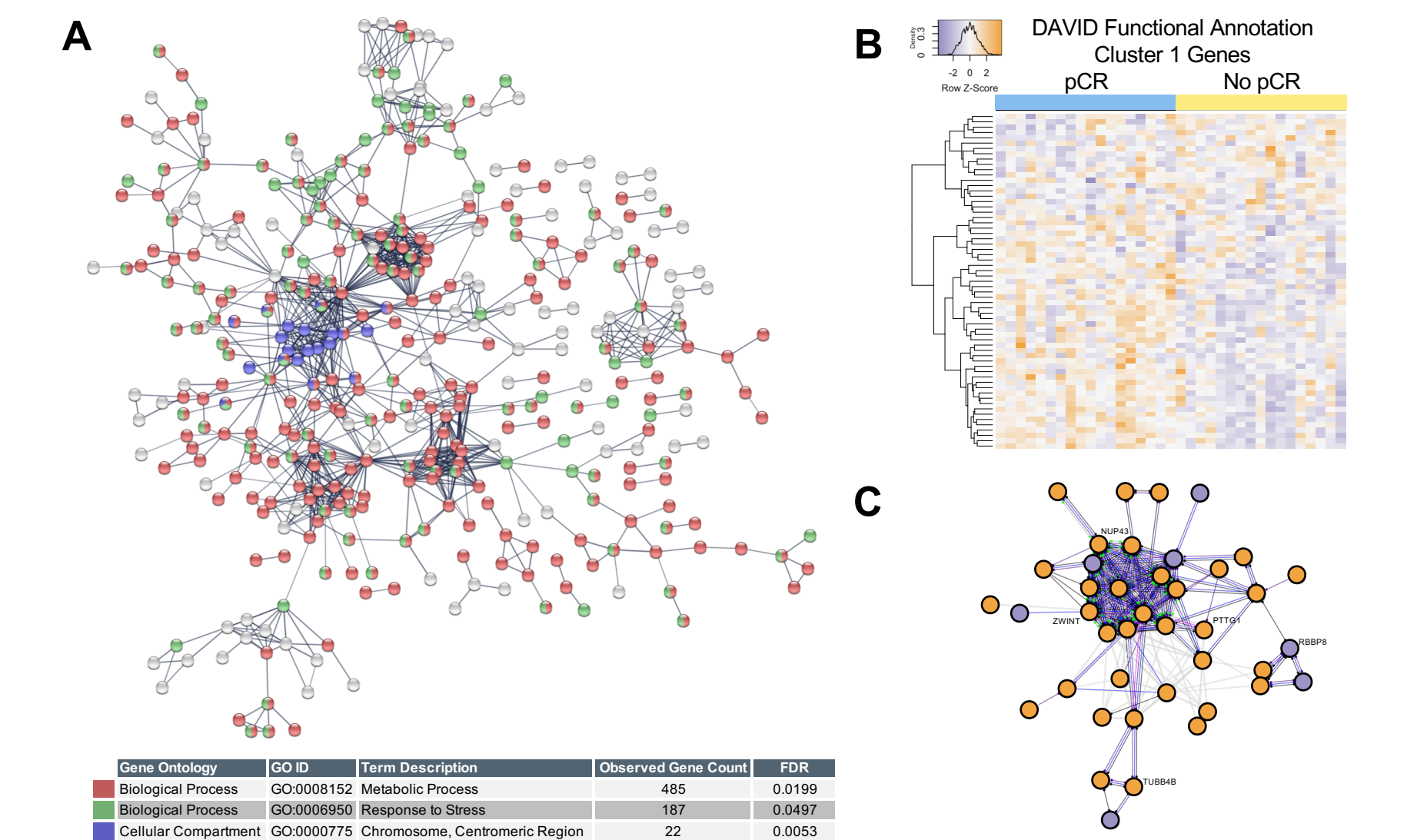


Figure 7: Case Study HR+HER2+ TDM1/P. (A) An unbiased approach discovers 902 ranked predictors, 867 are highly connected via protein-protein interaction (StringDB confidence > 0.7) and enrich for metabolic, stress response and cell cycle processes. (B-C) DAVID functional enrichment identifies a cluster of genes associated with cell cycle and mitosis that are upregulated (orange), e.g. NUP43, PTTG1, TUBB4B, ZWINT or downregulated (purple), e.g. RBBP8, in pCR vs. no pCR.

CONCLUSIONS

Our results suggests that hypothesis driven analysis restricted to assumed mechanisms of action of the experimental agents may be insufficient, and that exploration of possible off target effects may be needed to understand the underlying biology of response or resistance.

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